

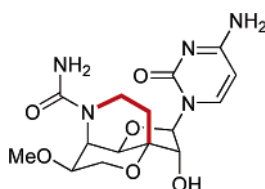
Structure-Based Organic Synthesis of a Tricyclic *N*-Malayamycin Analogue

Stephen Hanessian* and Dougal J. Ritson

Department of Chemistry, Université de Montréal, C. P. 6128, Succursale Centre-Ville, Montréal, Québec H3C 3J7, Canada

stephen.hanessian@umontreal.ca

Received September 14, 2006



The solid-state structure of crystalline malayamycin A reveals a urea substituent that bisects the plane of the chairlike tetrahydropyran subunit. On the basis of this topological feature, we synthesized a tricyclic *N*-nucleoside analogue in which an ethano bridge linked the urea NH group with the ring junction of the bicyclic tetrahydrofurofuran unit.

Introduction

Naturally occurring purine and pyrimidine nucleosides have been the cornerstones of the chemistry and biology of nature's genetic code since the beginning of creation.¹ *N*- and *C*-nucleosides with nontraditional heterocyclic as well as sugar components have also been found outside the DNA/RNA world.² Some of these have been endowed with impressive chemotherapeutic properties as anticancer, antiviral, and anti-infective agents in medical practice for decades.³

A select group of *N*- and *C*-pyrimidine nucleosides contains a bicyclic perhydrofurofuran "sugar" moiety rather than the

more commonly encountered monocyclic pentofuranosyl or hexopyranosyl residues. For example, ezomycin A₂⁴ **1** and octosyl acid A⁵ **2** are representatives of such bicyclic *N*-nucleosides, while ezomycin B₂⁶ **3** is a *C*-nucleoside equivalent (Figure 1). The ezomycins have been reported to exhibit antifungal and antibiotic activities.⁶ Quantamycin **4**, an unnatural synthetic analogue of lincomycin, was designed as a potential antibacterial agent.⁷ Some years ago, scientists at the Syngenta Crop Protection Laboratories in Jealott's Hill, U.K., isolated a new *C*-nucleoside from the soil organism *Streptomyces malaysiensis*, which they named malayamycin A (**5**).⁸ The gross structure of **5** was based on detailed NMR studies and degradation work. The proposed structure and stereochemical identity of **5** were recently confirmed by a total synthesis.⁹ Except for the commonly shared perhydrofurofuran core, the presence of a urea group, and the 5-substituted pyrimidinone units, the nature of functional groups and appendages in **5** were different from those in ezomycin B₂. Furthermore, **5** exhibited

(1) (a) Gesteland, T. R.; Cech, T. R.; Atkins, J. F. In *The RNA World*, 2nd ed.; Cold Springs Harbor Laboratory Press: New York, 1999. (b) Sarma, R. H.; Sarma, M. H. In *DNA Double Helix and the Chemistry of Cancer*; Adenine Press: New York, 1988. (c) Watson, J. D.; Crick, F. H. C. *Nature* **1953**, *171*, 737.

(2) For examples, see: (a) Ichikawa, S.; Kato, K. *Curr. Med. Chem.* **2001**, *8*, 3895. (b) Gao, H.; Mitra, A. K. *Synthesis* **2000**, 329. (c) Knapp, S. *Chem. Rev.* **1995**, *95*, 1859. (d) Postema, M. H. D. In *C-Glycoside Synthesis*; CRC Press: Boca Raton, FL, 1995. (e) Townsend, L. B. In *Chemistry of Nucleosides and Nucleotides*; Plenum Press: New York, 1994; pp 421–535.

(3) For examples, see: (a) Simons, C.; Wu, Q.; Htar, T. T. *Curr. Top. Med. Chem.* **2005**, *5*, 1191. (b) Rachakonda, S.; Cartee, L. *Curr. Med. Chem.* **2004**, *11*, 775. (c) Pathak, T. *Chem. Rev.* **2002**, *102*, 1623. (d) Hosmane, R. S. *Curr. Top. Med. Chem.* **2002**, *2*, 1093. (e) *Antisense Drug Technology*; Crooke, S. T., Ed.; Dekker: New York, 2001. (f) Mansur, T. S.; Storer, R. *Curr. Pharm. Des.* **1997**, *3*, 227.

(4) (a) Sakata, K.; Sakurai, A.; Tamura, S. *Agric. Biol. Chem.* **1975**, *39*, 885. (b) Sakata, K.; Sakurai, A.; Tamura, S. *Tetrahedron Lett.* **1974**, *49*, 4327. (c) Sakata, K.; Bakurai, A.; Tamura, S. *Agric. Biol. Chem.* **1973**, *39*, 697.

(5) For the total synthesis of octosyl acid, see: (a) Knapp, S.; Thakur, V. V.; Madurru, M. R.; Malolanarasimhan, K.; Morriello, G. J.; Doss, G. A. *Org. Lett.* **2006**, *8*, 1335. (b) Danishefsky, S.; Hungate, R. *J. Am. Chem. Soc.* **1986**, *108*, 2486. (c) Hanessian, S.; Kloss, J.; Sugawara, T. *J. Am. Chem. Soc.* **1986**, *108*, 2758. For isolation, see: (d) Isono, K.; Crain, P. K.; McCloskey, J. A. *J. Am. Chem. Soc.* **1975**, *97*, 943.

(6) Sakata, K.; Sakurai, A.; Tamura, S. *Tetrahedron Lett.* **1975**, 3191.
(7) Hanessian, S.; Sato, K.; Liak, T. J.; Danh, N.; Dixit, D.; Cheney, B. V. *J. Am. Chem. Soc.* **1984**, *106*, 6114.

(8) (a) Benner, J. P.; Boehlendorf, B. G. H.; Kipps, M. R.; Lamber, N. E. P.; Luck, R.; Molleyres, L.-P.; Neff, S.; Schuez, T. C.; Stanley, P. D. WO, 03/062242, CAN 139:132519. (b) Hanessian, S.; Machaalani, R.; Marcotte, S. WO, 04/069842.

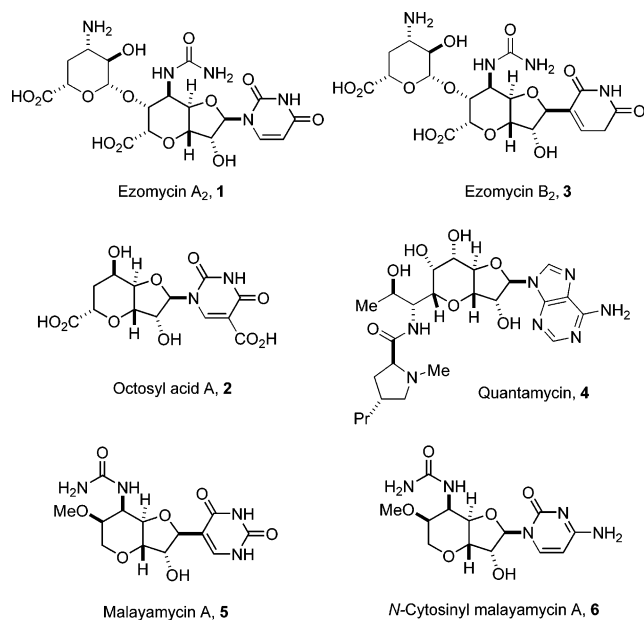


FIGURE 1. Natural (1, 2, 3, and 5) and unnatural (4 and 6) perhydrofuropranyl *N*- and *C*-nucleosides.

potent fungicidal activity,⁸ which instigated efforts toward the synthesis of *N*-purinyl and *N*-pyrimidinyl analogues.¹⁰ Indeed, a total synthesis of *N*-cytosinyl malayamycin A (6) revealed fungicidal activity at least equivalent to 5 (Figure 1).¹⁰

In the course of our synthetic efforts, we obtained X-ray quality crystals of 5 from water after slow evaporation. The three-dimensional solid-state structure as seen in the ORTEP diagram of one of the hydrated crystals revealed several interesting topological features (Figure 2). Most prominent was the orthogonal orientation of the axial C₅ urea group which bisects the plane of the chairlike tetrahydropyran ring of the bicyclic system, with the N–H group pointing “inward”. Previous functional group modifications¹⁰ have delineated the importance of stereochemistry as well as substitution to maintain fungicidal activity in this series. We therefore utilized the three-dimensional functional characteristics shown in the crystal structure of 5 to derive a tricyclic analogue 7 in which the axially oriented urea group was connected to C₃ (malayamycin A numbering) by an ethano bridge. Preliminary superposition of a modeled and minimized structure over the X-ray structure of 5 showed excellent congruence. Thus, we embarked on the synthesis of the proposed tricyclic analogue as part of a program dealing with structure-based organic synthesis.¹¹ In this approach, structural data available from bioactive natural products are used in the design and synthesis of unnatural congeners.¹²

Interestingly, such an approach was used many years ago in our *de novo* conception and synthesis of quantamycin 4, a hybrid structure intended to simulate recognition of the peptidyl amino acid transfer step by a modified lincomycin.^{7,13,14} The advent

(9) Hanessian, S.; Marcotte, S.; Machaalani, R.; Huang, G. *Org. Lett.* **2003**, *5*, 4277.

(10) (a) Loiseleur, O.; Schneider, H.; Huang, G. H.; Machaalani, R.; Selles, P.; Crowley, P.; Hanessian, S. *Org. Process Res. Dev.* **2006**, *10*, 518. (b) Hanessian, S.; Huang, G. H.; Chenel, C.; Machaalani, R.; Loiseleur, O. *J. Org. Chem.* **2005**, *70*, 6721. (c) Hanessian, S.; Marcotte, S.; Machaalani, R.; Huang, G. H.; Crowley, P. J.; Loiseleur, O. *WO*, 2005005432, 2005.

(11) Hanessian, S. *Chem. Med. Chem.*, published online Nov 8, 2006, <http://dx.doi.org/10.1002/cmdc.200600203>.

of X-ray crystal structures of complexes involving synthetic compounds with relevant enzymes has popularized so-called structure-based design in the quest for new therapeutic lead compounds.¹² Application of these notions to three-dimensional structural information in conjunction with bioactive conformations of natural products, alone or complexed with biological macromolecules such as enzymes¹⁵ and RNA,¹⁶ provides a powerful tool to explore synthetic chemistry in new directions.¹¹

Synthesis Plan

Clearly, a major challenge in the synthesis of 7 was the elaboration of the *N*–*C* ethano bridge in a stereocontrolled manner. In the disconnection illustrated in Scheme 1, we chose to first create the more demanding *C*-bridgehead tether and subsequently to engage it in aza-ring formation (Scheme 1). The allylic ether, readily available from diacetone-*D*-glucose, would be an appropriate substrate for a ring-closure metathesis reaction¹⁷ to generate the bicyclic core system. Following a stereo- and regiocontrolled functionalization of the double bond to give the *cis*-amino alcohol, the tether would be engaged in an intramolecular nucleophilic attack by the nitrogen to give the tricyclic core structure. Alternatively, the amino group in the tether could be the nucleophile. Elaboration of the acetal via anomeric activation, introduction of the cytosine, and functional group adjustments would afford the intended target 7.

(12) For related approaches, see: (a) *Structure-Based Drug Discovery: An Overview*; Hubbard, R. F., Ed.; Royal Society of Chemistry: Cambridge, 2006. (b) *Analogue-Based Drug Discovery*; Fischer, J., Ganellin, C. R., Eds.; Wiley-VCH: Weinheim, Germany, 2005. (c) Thiel, K. A. *Nat. Biotechnol.* **2004**, *22*, 513. (d) Anderson, A. C. *Chem. Biol.* **2003**, *10*, 787. (e) Klebe, G. *J. Mol. Med.* **2000**, *78*, 269. (f) Kubinyi, H. *Curr. Opin. Drug Discovery Dev.* **1998**, *1*, 4. (g) Bohacek, R. S.; McMartin, C.; Guida, W. C. *Med. Res. Rev.* **1996**, *16*, 3. See also: (h) Rees, D. C.; Congreve, M.; Murray, C. W.; Carr, R. *Nat. Rev. Drug Discovery* **2004**, *3*, 660. (i) Erlanson, D. A.; McDowell, R. S.; O'Brien, T. *J. Med. Chem.* **2004**, *47*, 3463. (j) Pellechia, M.; Sem, D. S.; Wuthrich, K. *Nat. Rev. Drug Discovery* **2002**, *1*, 211. (k) Fejzo, J.; Lepre, C. A.; Peng, J. W.; Bemis, G. W.; Murcko, M. A.; Moore, J. M. *Chem. Biol.* **1999**, *6*, 755. (l) Shuker, S. B.; Hajduk, P. J.; Meadows, R. P.; Fesik, S. W. *Science* **1996**, *274*, 1531. For an academic perspective, see: Hof, F.; Diederich, F. *J. Chem. Soc., Chem. Commun.* **2004**, 477.

(13) Cheney, B. V. *J. Med. Chem.* **1974**, *17*, 590.

(14) (a) Verdier, A. L.; Berthe, G.; Gharbi-Benarous, J.; Girauef, J.-P. *Bioorg. Med. Chem.* **2000**, *8*, 1225. (b) Fitzhugh, A. L. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 87.

(15) (a) Tyndall, J. D. A.; Nall, T.; Fairlie, D. P. *Chem. Rev.* **2005**, *105*, 973. (b) Loughlin, W. A.; Tyndall, J. D. A.; Glenn, M. P.; Fairlie, D. P. **2004**, *104*, 6085. (c) Greco, M. N.; Maryanoff, B. F. *Advances in Amino Acid Mimetics and Peptidomimetics*; JAI Press: Greenwich, CT, 2002; Vol. 1, p 41. (d) Babine, R. E.; Bender, S. L. *Chem. Rev.* **1997**, *97*, 1359. See also: (e) Sharff, A.; Jhota, H. *Curr. Opin. Chem. Biol.* **2003**, *7*, 340T. (f) Blundell, L.; Jhota, C.; Abell, C. *Nat. Rev. Drug Discovery* **2002**, *1*, 45. (g) Nienaber, V. L.; Richardson, P. L.; Klighofer, V.; Bouska, J. J.; Giranda, V. L.; Greer, J. *Nat. Biotechnol.* **2000**, *18*, 1105.

(16) (a) Ogle, J. M.; Ramakrishnan, V. *Annu. Rev. Biochem.* **2005**, *74*, 129. (b) Francois, B.; Russell, R. J. M.; Murray, J. B.; Aboulela, F.; Masquida, B.; Vicens, Q.; Westhof, E. *Nucleic Acids Res.* **2005**, *33*, 5677. (c) Vicens, Q.; Westhof, E. *J. Mol. Biol.* **2003**, *326*, 1175. (d) Hansen, J. L.; Moore, P. B.; Steitz, T. A. *J. Mol. Biol.* **2003**, *330*, 10611. (e) Vicens, Q.; Westhof, E. *Chem. Biol.* **2002**, *9*, 747. (f) Schlunzen, F.; Harms, J.; Tocilj, A.; Albrecht, R.; Yonath, A.; Franceschi, F. *Nature* **2001**, *413*, 814. (g) Vicens, Q.; Westhof, E. *Structure* **2001**, *9*, 647. (h) Wimberley, B. T.; Brodersen, D. E.; Clemons, W. M., Jr.; Morgan-Warren, R. J.; Caster, A. P.; Vonrhein, C.; Hartsch, T.; Ramakrishnan, V. *Nature* **2000**, *407*, 327. (i) Carter, A. P.; Clemons, W. M.; Brodersen, D. E.; Morgan-Warren, R. J.; Wimberley, B. T.; Ramakrishnan, V. *Nature* **2000**, *407*, 340. (j) Kirillov, S.; Porse, B. T.; Vester, B.; Woolley, P.; Garrett, R. A. *FEBS Lett.* **1997**, *406*, 223.

(17) For pertinent reviews, see: (a) Grubbs, R.; Chang, S. *Tetrahedron* **1998**, *54*, 4413. (b) Armstrong, S. K. *J. Chem. Soc., Perkin Trans.* **1998**, 371. (c) Furstner, A.; Picquet, M.; Bruneau, C.; Dixneuf, H. H. *J. Chem. Soc., Chem. Commun.* **1998**, 1315. (d) Schuster, M.; Blechert, S. *Angew. Chem., Int. Ed.* **1997**, *36*, 2036.

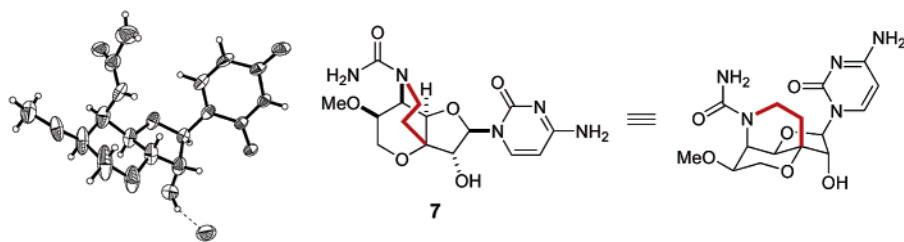
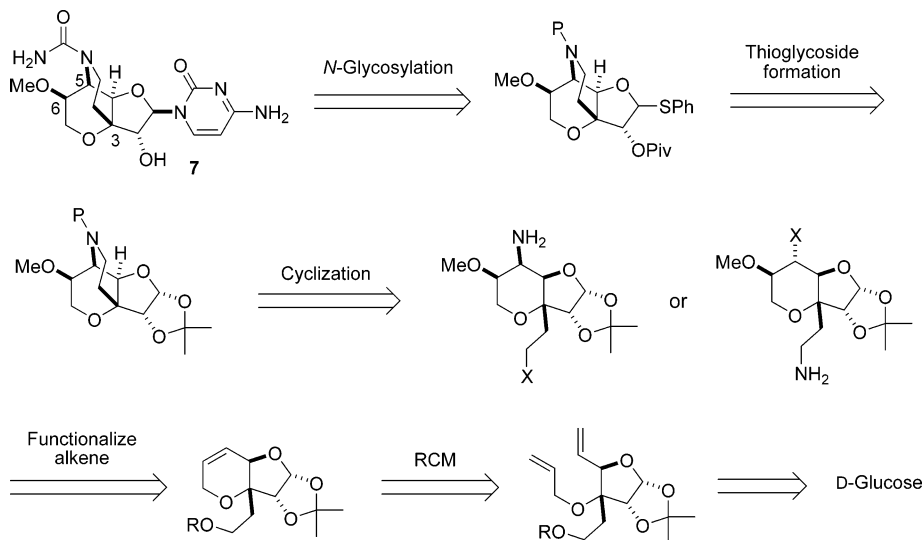


FIGURE 2. ORTEP diagram of malayamycin A hydrate and a tricyclic *N*-cytosinyl malayamycin A.

SCHEME 1. Disconnective Analysis



Although the essentials of this proposed route were previously accomplished in the synthesis of *N*-malayamycin A analogues,¹⁰ we were not in a position to ensure safe passage to **7**, being cognizant of the influence of steric effects and the uncertainty of ring strain in the elaboration of the acetal functionality in a bridged tricyclic ring system.

Results

Oxidation of diacetone-D-glucose **8** and treatment of the resulting ketone with allylmagnesium bromide gave the known¹⁸ *C*-allyl product **9** in 77% overall yield (Scheme 2). Ozonolysis followed by treatment of the ozonide with NaBH₄ in MeOH gave a cyclic alkoxyborane **10**, which necessitated treatment with ammonia in MeOH at 75 °C before the desired diol **11** could be liberated. Although *O*-benzylation of the primary hydroxyl group of **11** took place at room temperature in the presence of NaH in DMF, *O*-allylation of the tertiary hydroxyl group in **12** required heating at 100 °C in THF containing HMPA to give **13** in 97% yield. The distal acetone of **13** was selectively cleaved, and the diol **14**, after bis-mesylation, was subjected to a Finklestein-type elimination¹⁹ to give olefin **15**. Ring-closure metathesis employing Grubbs first-generation catalyst²⁰ led to the tethered bicyclic core **16** in 93% yield.

We then attempted to introduce the vicinal azido alcohol groups by first epoxidizing **16** to the α -epoxide **17** with *m*-CPBA, followed by opening with azide ion to the intended **18**. However, this sequence was abandoned because of the poor yield of epoxidation and the reluctance of the epoxide to open

with azide ion, no doubt due to a sterically impeded path caused by the benzyloxyethyl tether (Scheme 3). An alternative approach involved the *cis*-dihydroxylation of **16** in the presence of OsO₄ and NMO in aqueous acetone. The diol **19** (dr > 100:1) was easily separable from traces of the minor diastereoisomeric diol by column chromatography on silica gel. The configuration of **19** was established by nOe experiments. Mesylation of the diol **19** with 1.1 equiv of mesyl chloride at -78 °C afforded the monomesylate **20** selectively, most likely due to the pseudoequatorial disposition of the C₅ alcohol. The mesylate **20** was recovered unchanged when treated with NaN₃, even after prolonged periods of reflux in 2-methoxyethanol. Alternatively, treatment of **19** under Mitsunobu conditions with diphenylphosphoryl azide was also unsuccessful.

At this juncture, we reversed the order of bond-forming events leading to the desired aza-tricyclic system. Thus, diol **19** was debenzylated by catalytic hydrogenation, and the product was converted to the bis-tosylate **21** in 71% yield (Scheme 4). As expected, the pseudoaxial hydroxyl group at C₆ in **21** remained free after bis-tosylation. The position of the tosylate in **21** was confirmed via oxidation of the alcohol by the Dess–Martin periodinane reagent²¹ in CH₂Cl₂ to the corresponding ketone, and the latter compound was analyzed by ¹H NMR. Treatment of **21** with NaN₃ in DMF at 90 °C led to smooth and selective displacement of the primary tosylate to give **22** in 96% yield. We were now poised to effect an intramolecular ring closure

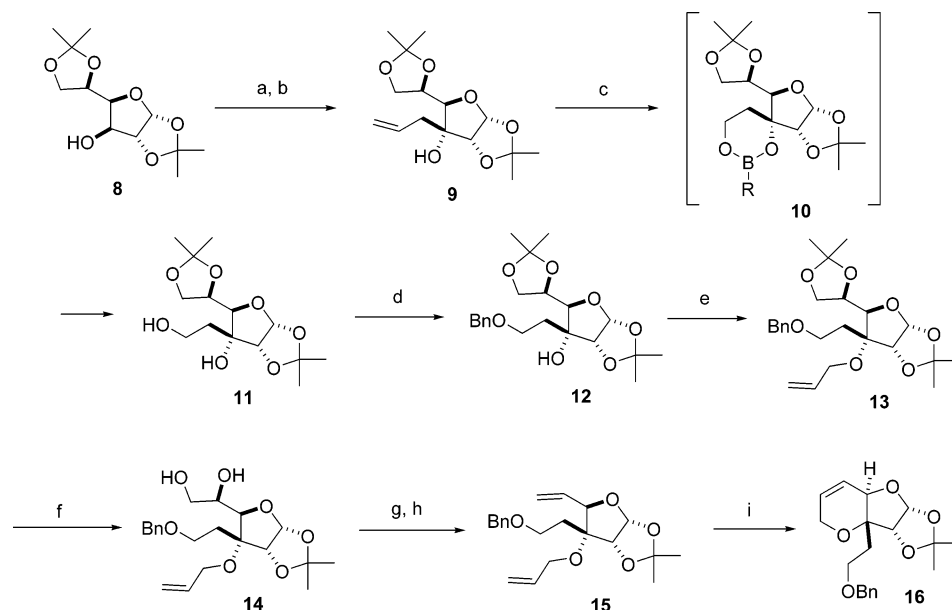
(20) Schwab, P.; Grubbs, R. H.; Ziller, J. W. *J. Am. Chem. Soc.* **1996**, *118*, 100.

(21) (a) Dess, D. B.; Martin, J. C. *J. Am. Chem. Soc.* **1991**, *113*, 7277. (b) Dess, D. B.; Martin, J. C. *J. Org. Chem.* **1983**, *48*, 4155. For a review, see: (c) Moriarty, R. M. *Org. React.* **1999**, *54*, 273.

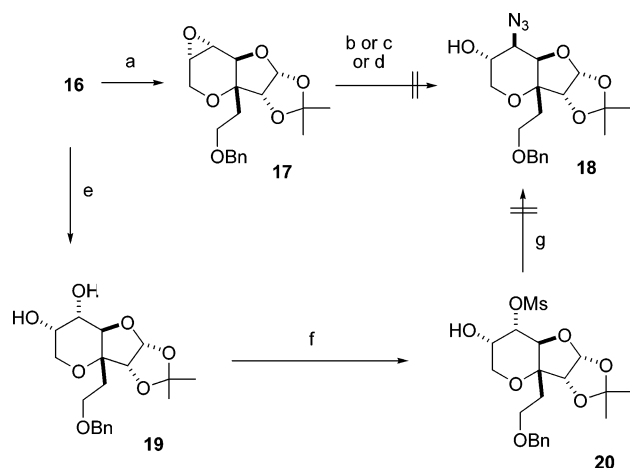
(18) Banerjee, S.; Ghosh, S. *J. Org. Chem.* **2003**, *68*, 3981.

(19) (a) Jones, J. K. N.; Thompson, J. L. *Can. J. Chem.* **1957**, *35*, 955.

(b) Finklestein, F. *Ber.* **1910**, *43*, 1528.

SCHEME 2. Synthesis of the Bicyclic Core^a

^a Reagents and conditions: (a) CrO₃, pyridine, Ac₂O, CH₂Cl₂, 0 °C to rt; (b) allylmagnesium bromide, THF, -10 °C to rt, 77% (two steps); (c) O₃, CH₂Cl₂, -78 °C then NaBH₄, MeOH, -78 °C to 75 °C, NH₃, rt to 75 °C then AcOH, rt, 59 to 84%; (d) NaH, BnBr, THF, 0 °C to rt, 88%; (e) NaH, allyl bromide, HMPA, THF, 100 °C, 97%; (f) 85% AcOH/H₂O, rt, 88%; (g) MsCl, Et₃N, CH₂Cl₂, 0 °C; (h) NaI, DMA, 100 °C, 83% (two steps); (i) 5 mol % Grubbs first-generation catalyst, CH₂Cl₂, rt, 93%.

SCHEME 3^a

^a Reagents and conditions: (a) *m*-CPBA, Na₂HPO₄, CH₂Cl₂, rt, 19%; (b) NaN₃, CH₂OHCH₂OMe, 130 °C; (c) NaN₃, NH₄Cl, MeCN/H₂O, 130 °C; (d) NaN₃, NH₄Cl, CH₂OHCH₂OMe, 130 °C; (e) 5 mol % OsO₄, NMO, acetone/H₂O, rt, 92%, dr > 100:1; (f) MsCl, Et₃N, CH₂Cl₂, -78 °C, 45%; (g) NaN₃, CH₂OHCH₂OMe, 155 °C.

from the azide extremity of the tether, after reduction to the primary amine, onto the C₅ tosylate group. In the event, reduction of the azide group in **22** in the presence of Pd black containing pyridine in EtOH, followed by heating in MeCN containing Et₃N, effected ring closure to the aza-tricyclic system **23**.

Before proceeding with the elaboration of the acetal into an anomerically activatable group, we had to select an appropriate *N*-protecting group that would be resistant to the acidic conditions required to cleave the 1,2-acetonide group and for the nucleobase coupling step, yet removable once the nucleosidic linkage was established. Initially, the 9-fluorenylmethyloxycarbonyl (Fmoc) protecting group was chosen. Addition of FmocCl

and pyridine to a solution of **23** in acetonitrile provided **24** in good overall yield. We were now in a position to “invert” the C₆ hydroxyl group and to *O*-methylate. To do so, we first oxidized the alcohol in **24** to the ketone **25** using the Dess–Martin periodinane reagent. Reduction with NaBH₄ in MeOH/CH₂Cl₂ was highly stereoselective giving the “inverted” alcohol **26** in good yield (the epimer was not observed). Being wary of forming a cyclic carbamate during *O*-methylation of **26** under basic conditions, we examined a number of acidic conditions for the *O*-methylation of **26**, such as CH₂N₂/silica gel,²² Me₂SO₄/NaHCO₃,²³ and Me₃O⁺BF₄⁻/proton sponge.²⁴ Unfortunately, all of these attempts were unsuccessful, and the use of MeI and Ag₂O in MeCN²⁵ resulted in the deprotection of the Fmoc group to give **27**. The trichloroethoxycarbonyl (Troc) group was then utilized due to its higher tolerance of basic conditions. However, exposure of **29**, which was synthesized in a manner analogous to that of **26**, to MeI and Ag₂O in acetonitrile led to the cyclic carbamate **30**.

It was clear that a far more robust protecting group would be required for the successful *O*-methylation of the C₆ alcohol. Accordingly, we opted for the *t*-butylsulfonyl (Bus) group, originally reported by Sun and Weinreb²⁶ and subsequently used in isolated instances only.²⁷ Treatment of **23** with *t*-butylsulfonyl chloride, followed by oxidation with *m*-CPBA, gave the desired *N*-Bus derivative **31** (Scheme 5). The oxidation–reduction sequence was repeated as before, and the alcohol **32** was

(22) Ohno, K.; Nishiyama, H.; Nagase, H. *Tetrahedron Lett.* **1979**, 4405.

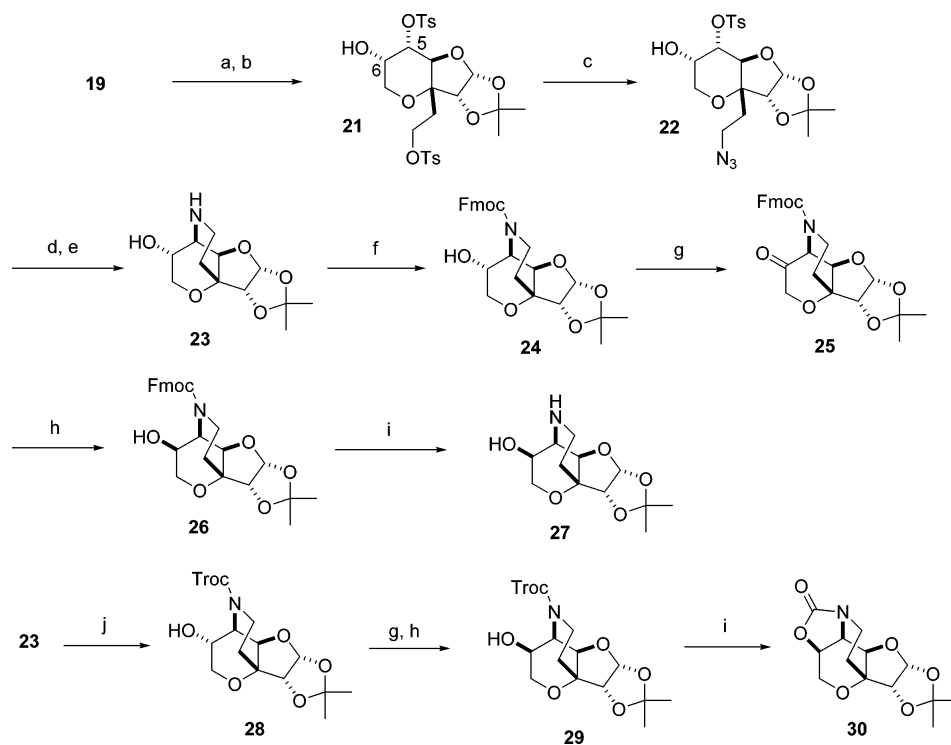
(23) Merz, A. *Angew. Chem., Int. Ed.* **1973**, 12, 846.

(24) Meerwein, H.; Hinz, G.; Hofmann, P.; Kroning, E.; Pfeil, E. *J. Prakt. Chem.* **1937**, 147, 257.

(25) Greene, A. E.; Drian, C. L.; Crabbe, P. *J. Am. Chem. Soc.* **1980**, 102, 7583.

(26) Sun, P.; Weinreb, S. M. *J. Org. Chem.* **1997**, 62, 8604.

(27) (a) Hanessian, S.; Del Valle, J. R.; Xue, Y.; Blomberg, N. *J. Am. Chem. Soc.* **2006**, 128, 10491. (b) Koep, S.; Gais, H.-J.; Raabe, G. *J. Am. Chem. Soc.* **2003**, 125, 13243. (c) Gontcharov, A. V.; Liu, H.; Sharpless, K. B. *Org. Lett.* **1999**, 1, 783.

SCHEME 4^a

^a Reagents and conditions: (a) Pd(OH)₂, H₂ (1 atm), EtOAc, rt; (b) TsCl, pyridine, -20 °C, 71% (two steps); (c) NaN₃, DMF, 90 °C, 96%; (d) Pd black, H₂ (1 atm), pyridine, EtOH, rt; (e) Et₃N, MeCN, 95 °C; (f) FmocCl, pyridine, MeCN, rt, 50% (three steps); (g) Dess–Martin periodinane, CH₂Cl₂, rt; (h) NaBH₄, MeOH/CH₂Cl₂, rt, 66% for **26**, 89% for **29** (two steps); (i) Ag₂O, MeI, MeCN, rt; (j) TrocCl, pyridine, MeCN, rt, 45% (three steps).

methylated under standard Williamson conditions to give the methyl ether **33**.

Previous reports from our laboratory^{7,9,10b,28} and the Knapp group²⁹ had shown the utility of γ -hydroxy dialkyl dithioacetals as intermediates for the synthesis of the alkyl thioglycosides, en route to nucleosidic bond formation in monocyclic as well as bicyclic systems. The feasibility of the same chemistry in the case of the bridged tricycle **33** as a precursor required some exploration. The mildest condition to effect acetal cleavage and formation of a diphenyldithioacetal was treatment of **33** with benzenethiol and Amberlyst-15 (H⁺) as a suspension in CH₂-Cl₂.¹⁰ The diphenyldithioacetal **34** was then treated with NBS in CH₂Cl₂ at 0 °C to effect cycloetherification,²⁸ affording **35** in 71% yield. A trace amount of a byproduct which may have been the β -anomer (not shown) was not isolated or characterized. Protection of the alcohol in **35** as the pivalate ester **36** gave X-ray quality crystalline material. As seen in the ORTEP diagram,³⁰ all the anticipated bond-forming sequences had taken place with the correct stereo- and regiochemistries. We continued the synthesis with the crystalline pivalate **36**, relying on its ability to direct anomeric substitution through neighboring group participation. The penultimate steps in the synthesis involved activation of the thioglycoside **36** with NIS/TfOH.^{29,31}

(28) Hanessian, S.; Dixit, D. M.; Liak, T. J. *Pure Appl. Chem.* **1981**, *53*, 129.

(29) (a) Knapp, S.; Shieh, W.-C.; Jaramillo, C.; Triller, R.; Nandau, S. R. *J. Org. Chem.* **1994**, *59*, 946. (b) Knapp, S.; Shieh, W.-C. *Tetrahedron Lett.* **1991**, *32*, 3627.

(30) For an ORTEP diagram of compound **36**, see Supporting Information S64.

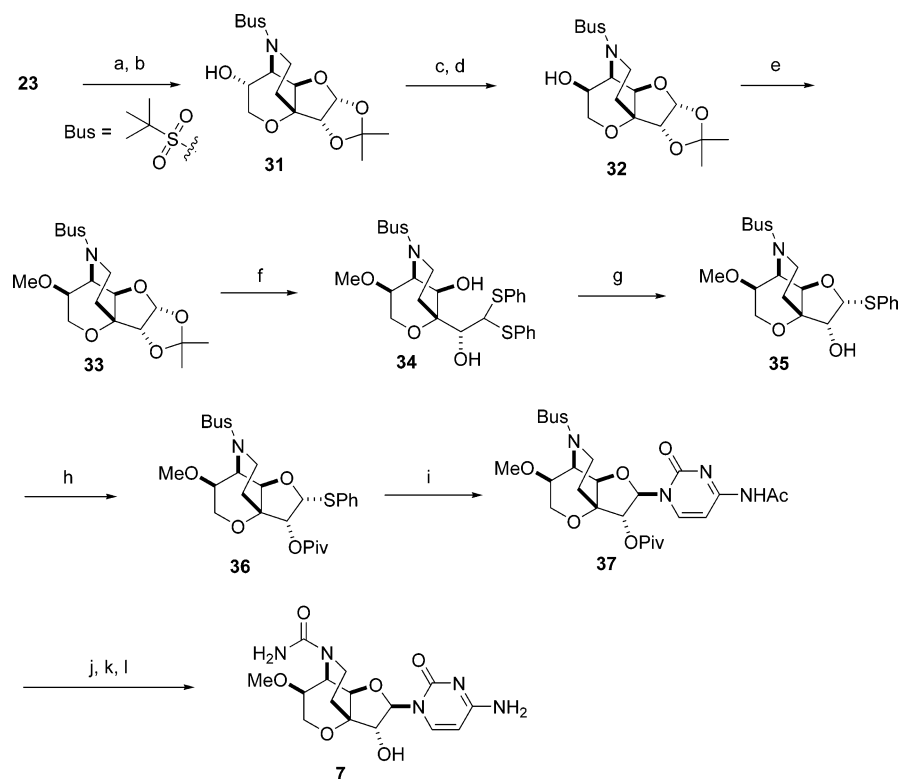
(31) Veeneman, G. H.; van Leeuwen, S. H.; van Boom, J. H. *Tetrahedron Lett.* **1990**, *31*, 1331.

and nucleophilic attack by the bis-TMS derivative of *N*-acetylcytosine to give **37** as a crystalline compound. Single-crystal X-ray analysis³² confirmed the 1,2-*trans*-disposition of the *N*-cytosinyl moiety. Finally, cleavage of the *N*-Bus group could be effected with TfOH in CH₂Cl₂ containing anisole. Subsequent installation of the urea group as previously reported^{9,10} and deprotection gave the intended tricyclic *N*-cytosinyl malayamycin A, **7**, as an amorphous, colorless solid. The synthesis of **7** was achieved starting with diacetone-D-glucose over 27 steps in an overall yield of 1.5%.

Discussion

The failure of nucleophilic attack by the azide ion, even under forcing conditions, of the epoxide **17** (or the mesylate **20**) is not surprising. It can be argued that the azide ion would have to approach the mesylate **20**, for example, from a trajectory that is sterically challenged. Furthermore, the required overlap with the σ^* C–O bond would be inaccessible in a chair conformation as in A (Figure 3). A boat conformer, B, would better expose the σ^* C–O orbital to the incoming charged nucleophile, although the benzyloxyethyl tether would still be an impediment. The intramolecular cyclization of the amine derived from **22** in MeCN at 90 °C diminishes the energetic penalty of a sterically impeded bimolecular attack by azide ion. A suprafacial trajectory of attack by the tethered aminoethyl group finds the requisite angle to effectively overlap with the σ^* orbital of the now axially disposed tosylate in **22**, presumably in a boat conformer, as shown in C (Figure 3).

(32) For an ORTEP diagram of compound **37**, see Supporting Information S73.

SCHEME 5^a

^a Reagents and conditions: (a) *t*-Butylsulfinyl chloride, Et₃N, rt; (b) *m*-CPBA, CH₂Cl₂, rt, 51% (four steps); (c) Dess–Martin periodinane, CH₂Cl₂, rt; (d) NaBH₄, CH₂Cl₂/MeOH, rt; (e) NaH, MeI, THF, rt, 98% (three steps); (f) PhSH, Amberlyst-15, CH₂Cl₂, rt, 80%; (g) NBS, CH₂Cl₂, 0 °C, 71%; (h) PivCl, DMAP, CH₂Cl₂/pyridine, rt, 81%; (i) 4-(*N*-trimethylsilyl)-acetamido-2-(trimethylsilyloxy)-pyrimidine, NIS, TFOH, CH₂Cl₂, rt, 52% (based on recovered starting material); (j) TFOH, anisole, CH₂Cl₂, rt; (k) Cl₃C(O)NCO, pyridine, rt; (l) MeNH₂, MeOH/H₂O, rt, 52% (three steps).

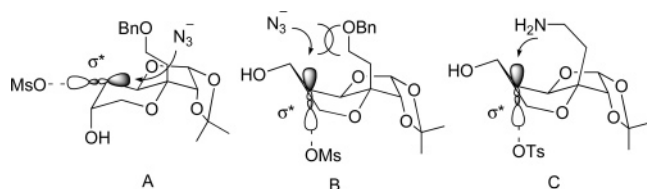


FIGURE 3. Inaccessible (A and B) approaches of an intermolecular nucleophile and the proposed boat conformation (C) allowing intramolecular cyclization.

Anomeric activation through oxocarbenium ion intermediates is at the core of glycoside and nucleoside chemistry.^{2,33} The ease of cycloetherification in going from the diphenyldithioacetal **34** to thioglycoside **35** deserves comment (Figure 4). This type of reactivity was at the basis of our early construction of nucleosides of perhydrofuroprans.²⁸ It was thought, however, that extension to a bridged, trans-fused dioxabicyclic precursor such as **35** would present additional torsional strain in the transition state involving intramolecular attack of the hydroxyl group onto the phenylthionium ion. This reaction is best done in the absence of an ester-protecting group next to the dithioacetal group because of its participating ability.^{10b} It is possible

that transient α -phenylthio epoxides may also be present (not shown). The successful cyclization to **35** in good yield is therefore remarkable.

Thioglycosides are well-known precursors to *O*-glycosides proceeding by activation with thiophilic reagents.³¹ In spite of precedents in the synthesis of bicyclic *N*-malayamycin A and related analogues,¹⁰ it is noteworthy that the formation of the nucleosidic bond in the tricycle **37** takes place in spite of the strained nature of the intermediates. Generation of the oxocarbenium ion A (Figure 4) must be followed by participation of the neighboring pivalate to the planar 1,2-dioxolenium ion B which imposes a fourth ring in the system. Attack of the pyrimidine base takes place in an anti-fashion to give the observed 1,2-trans-stereochemistry in **37**. Direct β -attack of the pyrimidine base on the oxocarbenium ion A is also possible. The apparent spatial tolerance of the axially disposed syn-aza-bridge with a bulky *N*-Bus group to the incoming *O*-silylated pyrimidine is certainly a felicitous result, in spite of the modest yield of this step.

Clearly, much remains to be learned from the chemistry of thionium and oxonium ions in these polycyclic systems in particular.^{33,34}

(33) See pertinent chapters in: (a) *The Organic Chemistry of Sugars*; Levy, D. E., Fugedi, P., Eds.; CRC Press: Boca Raton, FL, 2006. (b) *Carbohydrates in Chemistry and Biology*; Ernst, B., Hart, G. W., Sanaý, P., Eds.; Wiley-VCH: Weinheim, Germany, 2000. (c) *Preparative Carbohydrate Chemistry*; Hanessian, S., Ed.; Dekker: New York, 1997. (d) *Modern Methods in Carbohydrate Synthesis*; Khan, S. H., O'Neill, R. A., Eds.; Harwood Academic: Amsterdam, 1996.

(34) For insightful work on the reactivity and stereochemistry of oxocarbenium ions, see: (a) Shenoy, S. R.; Smith, D. M.; Woerpel, K. A. *J. Am. Chem. Soc.* **2006**, *128*, 8671. (b) Larsen, C. H.; Ridgway, B. H.; Shaw, J. T.; Smith, D. M.; Woerpel, K. A. *J. Am. Chem. Soc.* **2005**, *127*, 10879. (c) Ayala, L.; Lucero, C. G.; Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. *J. Am. Chem. Soc.* **2003**, *125*, 15521. (d) Schmitt, A.; Reissig, H.-U. *Eur. J. Org. Chem.* **2001**, 1169. (e) Schmitt, A.; Reissig, H.-U. *Eur. J. Org. Chem.* **2000**, 3893.

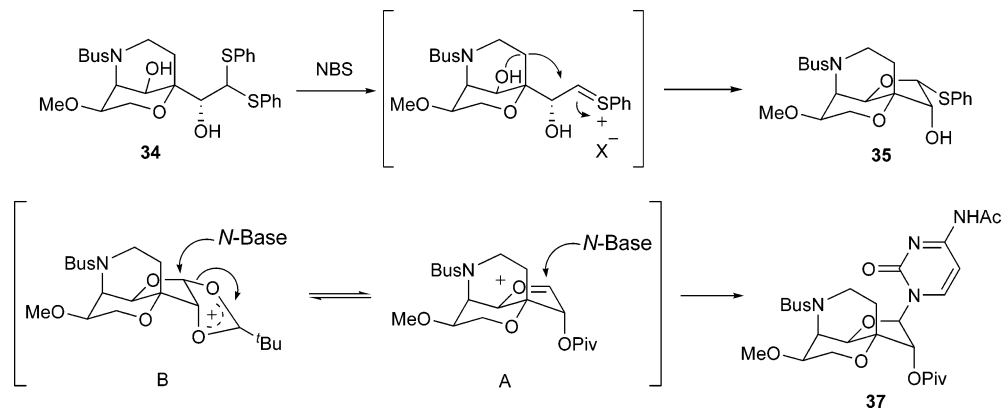


FIGURE 4. Activation at the anomeric center-thioglycoside and *N*-nucleoside formation.

Conclusion

We have conceived and synthesized a tricyclic analogue of *N*-malayamycin A in a stereocontrolled manner, based on the solid-state X-ray crystal structure of the parent malayamycin A. Topological information gleaned from the structure led to the choice of an ethano bridge tethering the proximal urea nitrogen atom with C₃ at the junction of the perhydrofurofuran ring system. Intramolecular cyclization from an amino terminal group on the tether was successfully performed onto a tosylate as a leaving group, possibly passing through a boatlike conformation to allow for better access to a σ^* orbital.

The utility of thionium and oxocarbenium ion intermediates in the construction of the tricyclic nucleoside highlights the successful completion of the synthesis. Unfortunately, preliminary testing of **7** as a fungicide did not show any activity, which may reflect a truly delicate balance between structure, function, and donor–acceptor interactions of the urea group in malayamycin A and its analogues in the presence of biological receptors and requisite enzymes. The possible role of the urea group in a preformed bioactive conformation of malayamycin A is presently under study.

Experimental Section

1,2:5,6-Di-*O*-isopropylidene-3-*C*-allyl-*D*-allofuranose (9**).** To a dry flask was charged CrO₃ (6.46 g, 64.6 mmol), under Ar atmosphere, containing anhydrous CH₂Cl₂ (110 mL), which was cooled to 0 °C. Anhydrous pyridine (11.8 mL, 11.5 g, 146 mmol) and Ac₂O (7.0 mL, 7.56 g, 74.0 mmol) were added followed by diacetone-*D*-glucose (10.0 g, 38.4 mmol), which was added portionwise over 30 min. After being stirred for 30 min, the reaction was brought to room temperature, and after a further 2 h the black solution was poured into EtOAc (ca. 400 mL). The mixture was filtered through silica washed with EtOAc. All the solvents were removed, and the residue was pumped overnight.

The crude oil was dissolved in anhydrous THF (120 mL) and added slowly to allylmagnesium bromide (1 M in Et₂O, 80 mL) at –10 °C. After the addition, the reaction was allowed to warm to room temperature and was stirred for 3 h. The reaction was then poured into ice/water (ca. 250 mL), and the majority of the solvent was evaporated in vacuo. The pH of the solution was taken to pH 7, with a solution of AcOH in Et₂O, and the products were extracted with Et₂O (3 × 200 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified by flash chromatography (hexanes/ethyl acetate, 85:15), which gave the product **9** (8.45 g, 28.2 mmol, 77%) as a colorless solid, and recrystallized from EtOAc/hexanes. *R*_f = 0.77 (hexanes/ethyl acetate, 1:1); mp 108–110; lit.¹⁸ 124 °C; [α]_D²⁵ +42.4 (*c* 1.40, CHCl₃); lit.¹⁸ [α]_D²⁵ +42.8 (*c* 3.0, CHCl₃); IR (neat)

ν 3474, 1374, 1073; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 6.01 (m, 1H), 5.69 (d, *J* = 3.7, 1H), 5.18 (m, 2H), 4.38 (d, *J* = 3.8, 1H), 4.16 (m, 2H), 3.93 (ddd, *J* = 9.3, 5.7, 4.2, 1H), 3.83 (d, *J* = 8.2, 1H), 2.67 (ddt, *J* = 14.4, 5.8, 1.5, 2H), 2.20 (dd, *J* = 14.5, 8.7, 1H), 1.61 (s, 3H), 1.48 (s, 3H), 1.39 (s, 3H), 1.36 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ (ppm) 132.5, 118.7, 112.3, 109.5, 103.4, 81.9, 81.1, 78.5, 73.0, 67.8, 36.6, 26.53, 26.52, 26.2, 25.1; LRMS (ESI) 301 (10%) [M + H]⁺.

3-*C*-(2-Hydroxyethyl)-1,2:5,6-di-*O*-isopropylidene-*D*-allofuranose (11**).** The alkene **9** (1.50 g, 5.00 mol) was dissolved in CH₂-Cl₂ (25 mL) and cooled to –78 °C. Ozone was bubbled through the solution until an excess was present. The excess ozone was removed by sparging O₂ through the solution, after which NaBH₄ (454 mg, 12.0 mmol) and MeOH (25 mL) were added. The reaction was brought to room temperature and then refluxed for 1.5 h. After being cooled to room temperature, NH_{3(g)} was bubbled through the solution for 30 min, followed by refluxing for a period of 8 h. The reaction was finally cooled and taken to pH 8 using MeOH/AcOH solution. The solvents were evaporated, and the material was purified by flash chromatography (7:3 to 1:9, hexanes/ethyl acetate) to provide **11** (1.27 g, 4.2 mmol, 84%) as a colorless solid. *R*_f = 0.13 (1:1, hexanes/ethyl acetate); mp 85 °C; [α]_D²⁵ +22.7 (*c* 0.67, CHCl₃); IR (neat) ν 3539, 3452, 1388; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 5.72 (d, *J* = 3.8, 1H), 4.56 (d, *J* = 3.9, 1H), 4.13 (m, 2H), 3.95 (m, 3H), 3.77 (d, *J* = 7.9, 1H), 2.75 (br s, 2H), 2.16 (ddd, *J* = 14.8, 8.1, 4.6, 1H), 1.61 (s, 3H), 1.58 (m, 1H), 1.46 (s, 3H), 1.38 (s, 6H); ¹³C NMR (75 MHz, CDCl₃) δ (ppm) 112.2, 109.3, 103.1, 81.9, 80.8, 79.4, 72.8, 67.4, 58.0, 32.6, 26.26, 26.27, 26.0, 24.9; HRMS (ESI) calcd for C₁₄H₂₄O₇Na [M + Na] 327.1414, found 327.1408

3-*C*-(2-Benzoyloxyethyl)-1,2:5,6-di-*O*-isopropylidene-*D*-allofuranose (12**).** A solution of **11** (5.34 g, 17.6 mmol) in anhydrous THF (65 mL) was cooled to 0 °C under an Ar atmosphere. NaH (60% dispersion in mineral oil, 850 mg, 21.3 mmol) was added portionwise and stirred for 1 h at 0 °C. BnBr (2.93 mL, 4.21 g, 24.6 mmol) was added dropwise, stirred for 30 min at 0 °C, then warmed to room temperature, and stirred for 3 days. A saturated solution of NH₄Cl (50 mL) was added slowly, and the layers were separated. The aqueous layer was extracted with Et₂O (3 × 60 mL), and the organic layers were combined, dried (Na₂SO₄), filtered, and concentrated. The residual material was purified by flash chromatography (85:15, hexanes/EtOAc), which yielded **12** (6.12 g, 15.5 mmol, 88%) as a colorless solid. *R*_f = 0.16 (4:1, hexanes/ethyl acetate); [α]_D²⁵ +19.7 (*c* 0.58, CHCl₃); IR (neat) ν 3480, 1386, 1373; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.34 (m, 5H), 5.70 (d, *J* = 3.7, 1H), 4.69 (d, *J* = 3.7, 1H), 4.56 (d, *J* = 12.7, 1H), 4.52 (d, *J* = 12.6, 1H), 4.13 (m, 2H), 3.92 (dd, *J* = 11.4, 8.4, 1H), 3.85 (m, 2H), 3.76 (dt, *J* = 9.4, 5.7, 1H), 2.89 (br s, 1H), 2.14 (ddd, *J* = 14.6, 5.6, 5.6, 1H), 1.78 (ddd, *J* = 14.6, 7.4, 5.6, 1H), 1.61 (s, 3H), 1.47 (s, 3H), 1.37 (s, 3H), 1.36 (s, 3H); ¹³C NMR (75 MHz,

CDCl₃) δ (ppm) 137.7, 128.1, 127.4, 127.7, 112.1, 109.3, 103.2, 81.8, 81.0, 78.6, 72.93, 72.90, 67.6, 65.4, 31.1, 26.35, 26.33, 26.0, 24.9; HRMS (ESI) calcd for C₂₁H₃₀O₇Na [M + Na] 417.1884, found 417.1882.

3-*O*-Allyl-3-*C*-(2-benzyloxyethyl)-1,2:5,6-di-*O*-isopropylidene-D-allofuranose (13). To a solution of **12** (7.75 g 19.7 mmol) in anhydrous THF (110 mL), under Ar atmosphere, was added NaH (60% dispersion, 1.25 g, 31.3 mmol) portionwise. The reaction was heated to gentle reflux for 2 h and then cooled to room temperature when HMPA (12 mL) was added, followed by allyl bromide (3.60 mL, 4.98 g, 41.2 mmol). The reaction was refluxed at 95 °C for 1.5 h and cooled to room temperature, and H₂O (100 mL) was added. The products were extracted with Et₂O (3 × 100 mL), dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by flash chromatography (4:1, hexanes/ethyl acetate) to provide **13** (8.33 g, 19.1 mmol, 97%) as pale yellow oil. $R_f = 0.25$ (4:1, hexanes/ethyl acetate); [α]_D²⁵ +41.0 (*c* 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.36 (m, 5H), 5.93 (ddt, $J = 17.2$, 10.4, 5.2, 1H), 5.62 (d, $J = 3.5$, 1H), 5.30 (dtd, $J = 17.2$, 3.6, 1.7, 1H), 5.13 (dtd, $J = 10.4$, 3.4, 1.5, 1H), 4.62 (d, $J = 3.5$, 1H), 4.52 (s, 2H), 4.32 (dt, $J = 5.2$, 1.6, 1H), 4.19 (dt, $J = 5.1$, 1.6, 1H), 4.04–4.17 (m, 3H), 3.92 (dd, $J = 8.0$, 5.6, 1H), 3.74 (m, 2H), 2.18 (dt, $J = 14.8$, 6.6, 1H), 1.89 (dt, $J = 14.8$, 6.9, 1H), 1.59 (s, 3H), 1.44 (s, 3H), 1.35 (s, 3H), 1.34 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 137.8, 135.0, 128.1, 127.34, 127.29, 115.2, 112.3, 109.2, 102.8, 83.2, 83.1, 80.9, 72.8, 72.6, 68.1, 65.8, 65.2, 30.4, 26.6, 26.13, 26.12, 25.0; HRMS (ESI) calcd for C₂₄H₃₄O₇Na [M + Na] 457.2197, found 457.2194.

3-*O*-Allyl-3-*C*-(2-benzyloxyethyl)-1,2-*O*-isopropylidene-D-allofuranose (14). Compound **13** (9.91 g, 22.8 mmol) was stirred in AcOH/H₂O (85:15 v/v, 100 mL) for 48 h at room temperature. A saturated solution of NaHCO₃ (200 mL) was added slowly, and the reaction was neutralized carefully with solid NaHCO₃. The products were extracted with Et₂O (3 × 250 mL), dried (Na₂SO₄), filtered, and concentrated, and the crude material was purified by flash chromatography (7:3 to 1:1, hexanes/ethyl acetate), which gave **14** (7.90 g, 20.1 mmol, 88%) as a pale yellow oil. $R_f = 0.36$ (1:1, hexanes/ethyl acetate); [α]_D²⁵ +47.6 (*c* 0.53, CHCl₃); IR (neat) ν 3436; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.37 (m, 5H), 5.90 (m, 1H), 5.66 (d, $J = 3.6$, 1H), 5.24 (ddt, $J = 17.2$, 3.3, 1.6, 1H), 5.14 (ddt, $J = 10.5$, 3.0, 1.4, 1H), 4.59 (d, $J = 3.6$, 1H), 4.53 (d, $J = 11.8$, 1H), 4.50 (d, $J = 11.8$, 1H), 4.25 (ddt, $J = 12.1$, 5.0, 1.6, 1H), 4.15 (ddt, $J = 12.1$, 5.5, 1.4, 1H), 4.07 (d, $J = 9.0$, 1H), 3.85 (m, 2H), 3.69 (m, 3H), 2.90 (br s, 1H), 2.20 (dt, $J = 15.0$, 6.9, 1H), 1.89 (dt, $J = 14.8$, 6.9, 1H), 1.80 (br s, 1H), 1.59 (s, 3H), 1.34 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 137.4, 134.2, 128.1, 127.5, 127.4, 116.1, 112.5, 103.6, 84.0, 82.1, 78.3, 72.9, 69.3, 66.5, 65.1, 64.3, 30.9, 26.4, 26.1; LRMS (ESI) 395 (25%) [M + H]⁺.

(3aR,5S,6R,6aR)-6-Allyloxy-6-(2-benzyloxyethyl)-2,2-dimethyl-5-vinyltetrahydrofuro[2,3-*d*][1,3]dioxole (15). A solution of **14** (7.90 g, 20.1 mmol) in anhydrous CH₂Cl₂ (110 mL), under an Ar atmosphere, was cooled to 0 °C. Et₃N (7.10 mL, 5.13 g, 50.3 mmol) was added, and the temperature of the solution was allowed to equilibrate before MsCl (3.40 mL, 5.03 g, 43.9 mmol) was added dropwise. The reaction was stirred at 0 °C for 1 h, then H₂O (55 mL) was added, and the reaction was stirred for a further 10 min at this temperature. The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 50 mL). The organics were combined, dried (Na₂SO₄), filtered, and concentrated.

The oily residue was dissolved in anhydrous dimethylacetamide (110 mL), and NaI (19.6 g, 131 mmol) was added. The reaction was heated to 100 °C for 6 h and then cooled to room temperature, and a saturated solution of Na₂S₂O₃ (100 mL) was poured into the reaction mixture. Stirring was continued until the color had disappeared, then H₂O (100 mL) was added, and the products were extracted with Et₂O (3 × 200 mL), dried (Na₂SO₄), and filtered. After evaporation of the solvents in vacuo, the residue was purified by flash chromatography (9:1, hexanes/ethyl acetate), which gave

15 (5.97 g, 16.6 mmol, 83%) as a very pale yellow oil. $R_f = 0.91$ (1:1, hexanes/ethyl acetate); [α]_D²⁵ +41.2 (*c* 0.49, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.34 (m, 5H), 5.91 (ddt, $J = 17.2$, 10.4, 5.3, 1H), 5.82 (ddd, $J = 16.7$, 10.7, 5.8, 1H), 5.72 (d, $J = 3.7$, 1H), 5.46 (dt, $J = 17.3$, 1.7, 1H), 5.28 (m, 2H), 5.15 (ddd, $J = 10.4$, 3.2, 1.7, 1H), 4.64 (m, 1H), 4.58 (d, $J = 3.7$, 1H), 4.50 (s, 2H), 4.14 (dt, $J = 5.4$, 1.5, 2H), 3.64 (dt, $J = 6.8$, 1.2, 2H), 2.07 (dt, $J = 14.8$, 6.8, 1H), 1.80 (dt, $J = 14.9$, 6.7, 1H), 1.60 (s, 3H), 1.34 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 137.8, 134.7, 132.1, 128.1, 127.32, 127.26, 118.2, 115.8, 112.2, 103.1, 83.7, 82.3, 81.1, 72.7, 65.5, 65.1, 30.2, 26.5, 26.1; HRMS (ESI) calcd for C₂₁H₂₈O₅Na [M + Na] 383.1829, found 383.1819.

(3aR,3bS,7aS,8aR)-3b-(2-Benzyloxyethyl)-2,2-dimethyl-3a,5,7a,8a-tetrahydro-3bH-1,3,4,8-tetraoxacyclopenta[*a*]indene (16). A solution of **15** (5.97 g, 16.6 mmol) in anhydrous CH₂Cl₂ (2.1 L) was degassed, and an Ar atmosphere was applied. Grubbs first-generation catalyst (4 mol %, 0.66 mmol, 542 mg) was added, and the reaction was stirred overnight at room temperature. EtOAc (100 mL) was then added, and the reaction was filtered through silica washing with CH₂Cl₂/EtOAc (9:1). The solvents were removed, and the oily residue was purified by flash chromatography (4:1, hexanes/ethyl acetate) to yield **16** (5.18 g, 15.6 mmol, 93%) as a viscous, pale brown oil. $R_f = 0.74$ (1:1, hexanes/ethyl acetate); [α]_D²⁵ -5.6 (*c* 0.87, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.33 (m, 5H), 6.18 (m, 1H), 5.83 (d, $J = 3.4$, 1H), 5.66 (ddd, $J = 10.5$, 5.5, 2.5, 1H), 4.82 (dd, $J = 3.5$, 0.8, 1H), 4.68 (m, 1H), 4.57 (d, $J = 11.8$, 1H), 4.52 (d, $J = 11.8$, 1H), 4.42 (ddd, $J = 17.9$, 5.4, 2.6, 1H), 4.22 (ddt, $J = 17.9$, 3.7, 2.5, 1H), 3.78 (ddd, $J = 9.6$, 6.9, 5.8, 1H), 3.68 (m, 1H), 2.18 (dt, $J = 15.6$, 5.5, 1H), 1.62 (s, 3H), 1.44 (ddd, $J = 15.6$, 7.9, 6.2, 1H), 1.36 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 137.9, 128.0, 127.2, 125.7, 123.9, 112.6, 105.6, 79.0, 78.4, 77.4, 72.8, 72.5, 65.5, 64.5, 27.4, 25.8, 25.5; HRMS (ESI) calcd for C₁₉H₂₄O₅Na [M + Na] 355.1516, found 355.1507.

(3aR,3bR,6S,7S,7aS,8aR)-3b-(2-Benzyloxyethyl)-2,2-dimethylhexahydro-1,3,4,8-tetraoxacyclopenta[*a*]indene-6,7-diol (19). To a solution of **16** (5.18 g, 15.6 mmol) in acetone/H₂O (8:1, 130 mL) were added 4-methylmorpholine *N*-oxide (3.67 g, 31.3 mmol) and then 2.5 wt % solution of OsO₄ in *t*-BuOH (9.78 mL, ca. 3.5 mol %) at room temperature. After 2.5 h, a saturated solution of Na₂S₂O₃ (100 mL) was added, and the solution was stirred overnight. The majority of the acetone was removed in vacuo, and the black solution was extracted with EtOAc (3 × 100 mL), dried (MgSO₄), filtered, and concentrated. The residue was purified by flash chromatography (9:1, ethyl acetate/hexanes) to yield **19** (5.28 g, 14.4 mmol, 92%) as an off-white foam and >100:1 mixture of diastereoisomers, the minor, of which, was not characterized. $R_f = 0.33$ (ethyl acetate); [α]_D²⁵ +59.7 (*c* 1.7, CHCl₃); IR (neat) ν 3446, 2250; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.36 (m, 5H), 5.79 (d, $J = 3.4$, 1H), 4.69 (d, $J = 3.5$, 1H), 4.56 (d, $J = 11.7$, 1H), 4.51 (d, $J = 11.7$, 1H), 4.23 (d, $J = 10.5$, 1H), 4.11 (m, 2H), 4.01 (m, 1H), 3.85 (dd, $J = 13.5$, 2.0, 1H), 3.72 (dt, $J = 9.5$, 6.9, 1H), 3.64 (ddd, $J = 9.6$, 7.1, 5.4, 1H), 2.50 (br s, 1H), 2.20 (dt, $J = 15.5$, 5.4, 1H), 1.85 (br s, 1H), 1.63 (s, 3H), 1.47 (dt, $J = 15.5$, 7.1, 1H), 1.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 137.6, 128.1, 127.4, 127.3, 113.3, 104.4, 80.7, 79.7, 75.1, 73.0, 68.6, 67.7, 66.8, 64.6, 25.8, 25.7, 25.4; HRMS (ESI) calcd for C₁₉H₂₇O₇ [M + H] 367.1751, found 367.1768.

Bis-*p*-toluenesulfonate Ester 21. Compound **19** (364 mg, 0.99 mmol) was dissolved in EtOAc (10 mL), and Pd(OH)₂/C 20 wt % (40 mg) was added. The suspension was degassed, and an H₂ atmosphere was applied (1 atm). The reaction was stirred at room temperature for 2 h, and then the suspension was filtered and washed with hot EtOAc and then warm MeOH. The solution was concentrated in vacuo to provide a colorless solid.

The crude material was dissolved in anhydrous pyridine (9 mL) and cooled to -20 °C. TsCl (850 mg, 4.46 mmol) was added in three portions with 3-h intervals, and the reaction was stirred at -20 °C overnight. MeOH was then added, the reaction was stirred

for 10 min, and the solvents were removed in vacuo. The residue was dissolved in H₂O (20 mL) and extracted with Et₂O (3 × 20 mL), dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by flash chromatography (3:2 to 1:1, hexanes/ethyl acetate) to yield **21** (414 mg, 0.71 mmol, 71%) as a colorless foam. $R_f = 0.22$ (1:1, hexanes/ethyl acetate); $[\alpha]_D^{25} +32.1$ (c 0.43, CHCl₃); IR (neat) ν 3525, 1598, 1358, 1176; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.84 (d, $J = 8.4$, 2H), 7.80 (d, $J = 8.4$, 2H), 7.39 (d, $J = 7.9$, 2H), 7.37 (d, $J = 7.9$, 2H), 5.64 (d, $J = 3.5$, 1H), 4.79 (dd, $J = 11.5$, 3.5, 1H), 4.39 (d, $J = 11.5$, 1H), 4.37 (d, $J = 3.5$, 1H), 4.28 (m, 1H), 4.15 (m, 2H), 4.08 (dd, $J = 13.6$, 1.3, 1H), 3.70 (dd, $J = 13.5$, 1.9, 1H), 2.54 (br s, 1H), 2.49 (s, 6H), 2.23 (m, 1H), 1.58 (s, 3H), 1.54 (m, 1H), 1.31 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 145.1, 145.0, 132.5, 132.2, 129.7, 129.6, 127.6, 127.5, 113.7, 104.1, 81.3, 79.1, 77.5, 71.4, 67.6, 66.7, 64.6, 25.7, 25.6, 25.1, 21.4, 21.3; LRMS (ESI) 585 (100%) [M + H]⁺.

Azide 22. Compound **21** (414 mg, 0.71 mmol) was dissolved in anhydrous DMF (10 mL), and NaN₃ (46 mg, 0.71 mmol) was added before the reaction was heated to 90 °C. After 50 min, LC–MS indicated consumption of starting material, and therefore the solution was cooled to room temperature and diluted with brine (15 mL). The product was extracted with EtOAc (3 × 15 mL), and the combined organics were washed with H₂O (2 × 10 mL), then dried (MgSO₄), filtered, and concentrated. The crude material was purified by flash chromatography (1:1, hexanes/ethyl acetate) to yield **22** (311 mg, 0.68 mmol, 96%) as a colorless foam. $R_f = 0.34$ (1:1, hexanes/ethyl acetate); $[\alpha]_D^{25} +43.5$ (c 0.95, CHCl₃); IR (neat) ν 3503, 2102; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.86 (d, $J = 8.3$, 2H), 7.38 (d, $J = 8.0$, 2H), 5.70 (d, $J = 3.5$, 1H), 4.89 (dd, $J = 11.5$, 3.5, 1H), 4.53 (d, $J = 3.5$, 1H), 4.43 (d, $J = 11.6$, 1H), 4.33 (m, 1H), 4.14 (dd, $J = 13.5$, 1.3, 1H), 3.81 (dd, $J = 13.5$, 2.0, 1H), 3.47 (m, 2H), 2.48 (s, 3H), 2.10 (m, 2H), 1.60 (s, 3H), 1.33 (s, 3H), 1.32 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 145.0, 132.6, 129.5, 127.6, 113.7, 104.2, 81.5, 79.2, 77.6, 71.5, 67.8, 66.6, 45.2, 25.8, 25.7, 24.9, 21.4; HRMS (ESI) calcd for C₁₉H₂₅N₃O₈-SNa [M + Na] 478.1255, found 478.1247.

***N*-9-Fluorenylmethylcarbamate 24.** To a solution of **22** (100 mg, 0.22 mmol) in EtOH/pyridine (99:1, 5.5 mL) was added Pd black (16 mg), and the suspension was degassed, and an H₂ atmosphere (1 atm) was applied. After being stirred at room temperature for 1.5 h, the suspension was filtered and washed with warm MeOH/pyridine (99:1), and the solution was concentrated and dried on the pump overnight.

The crude material was dissolved in MeCN (6 mL) with Et₃N (90 μ L, 66 mg, 0.66 mmol), and the reaction was refluxed (95 °C) for 24 h. The solution was then cooled to 0 °C, and pyridine (18 μ L, 17 mg, 0.22 mmol) and FmocCl (63 mg, 0.24 mmol) were added. After 3 h, LC–MS indicated the consumption of the amine, and the solvents were removed in vacuo. The brown residue was dissolved in 0.2 M HCl (5 mL), and the products were extracted with EtOAc (3 × 15 mL), dried (MgSO₄), filtered, and concentrated. The crude material was purified by flash chromatography (1:1 to 7:3, ethyl acetate/hexanes) to yield **24** (54 mg, 0.11 mmol, 50%) as a colorless foam. $R_f = 0.16$ (1:1, ethyl acetate/hexanes); $[\alpha]_D^{25} 10.6$ (c 1.8, CHCl₃); IR (neat) ν 3449, 1697; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.78 (m, 2H), 7.58 (m, 2H), 7.30–7.46 (m, 4H), 5.76 (d, $J = 3.3$, 0.4H), 5.72 (d, $J = 3.3$, 0.6H), 4.67 (dd, $J = 10.7$, 5.2, 0.4H), 4.63 (dd, $J = 10.7$, 4.7, 0.6H), 4.55 (m, 1.4H), 4.45 (dd, $J = 10.8$, 6.2, 0.4H), 4.33 (d, $J = 3.1$, 0.6H), 4.31 (d, $J = 0.4$ H), 4.25 (m, 1.6H), 4.23 (m, 1H), 3.90 (m, 1.4H), 3.70 (d, $J = 13.6$, 0.6H), 3.43 (m, 2H), 3.28 (br s, 0.6H), 2.75 (m, 1H), 1.64 (s, 1.2H), 1.61 (s, 1.8H), 1.38 (s, 1.2H), 1.35 (s, 1.8H); ¹³C NMR (75 MHz, CDCl₃) δ (ppm) 149.1, 148.6, 137.1, 138.0, 137.9, 137.7, 135.8, 135.58, 135.54, 135.4, 123.2, 123.0, 122.8, 122.6, 122.45, 122.40, 120.6, 120.4, 120.3, 115.88, 115.85, 115.80, 115.7, 110.7, 110.6, 101.6, 81.9, 76.6, 73.5, 73.4, 70.0, 68.4, 68.9, 67.5, 66.8, 56.4, 54.4, 54.1, 49.1, 49.0, 40.0, 39.8, 31.2, 31.0, 29.8, 29.5; HRMS (ESI) calcd for C₂₇H₃₀NO₇ [M + H] 480.2017, found 480.2020.

Alcohol 26. To a solution of **24** (26 mg, 0.054 mmol) in dry

CH₂Cl₂ (1 mL) was added Dess–Martin periodinane (34 mg, 0.081 mmol) at room temperature. After 2.5 h, a saturated solution of Na₂S₂O₃ (2 mL) was added, and the reaction was stirred for 30 min. The products were extracted with CH₂Cl₂ (3 × 3 mL), and the combined organics were washed with a saturated solution of NaHCO₃ (2 × 4 mL), dried (Na₂SO₄), filtered, and concentrated.

The residue was dissolved in CH₂Cl₂/MeOH (1:1, 1 mL) at room temperature, and to the stirred solution was added NaBH₄ (4 mg, 0.11 mmol). After 30 min, a saturated solution of NH₄Cl (2 mL) was added, and the reaction was stirred for 10 min. The layers were separated. The aqueous layer was extracted with EtOAc (3 × 4 mL), after which the organics were combined, dried (MgSO₄), and filtered, and the solvent was removed. The solid residue was purified by flash chromatography (3:2, ethyl acetate/hexanes) to yield **26** (17 mg, 0.035 mmol, 66%) as a colorless solid. $R_f = 0.19$ (1:1, ethyl acetate/hexanes); mp 90–93 °C; $[\alpha]_D^{25} -3.4$ (c 0.85, CHCl₃); IR (neat) ν 3432, 1694; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.79 (d, $J = 7.6$, 2H), 7.59 (d, $J = 7.4$, 2H), 7.44 (t, $J = 7.3$, 2H), 7.34 (t, $J = 7.3$, 2H), 5.81 (d, $J = 3.0$, 0.7H), 5.77 (br s, 0.3H), 4.91 (s, 0.7H), 4.55 (m, 1.3H), 4.43 (dd, $J = 10.4$, 6.7, 0.7H), 4.34 (d, $J = 2.9$, 0.7H), 4.27 (t, $J = 6.5$, 1H), 4.15 (br s, 0.3H), 3.85–4.25 (m, 3.7H), 3.80 (br s, 0.3H), 3.53–3.74 (m, 2.0H), 3.44 (m, 0.3H), 2.20 (m, 0.7H), 1.95 (m, 0.3H), 1.63 (m, 4H), 1.39 (s, 2.1H), 1.35 (s, 0.9H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 158.8, 143.3, 143.2, 141.0, 127.5, 127.0, 126.8, 124.7, 124.5, 124.2, 124.1, 119.7, 114.2, 105.2, 82.1, 81.3, 70.8, 76.9, 73.1, 72.2, 68.5, 67.8, 67.3, 66.8, 64.4, 52.8, 46.8, 38.8, 37.5, 28.4, 26.8, 26.2, 26.0, 25.7; HRMS (ESI) calcd for C₁₆H₂₈NO₇S [M + H] 378.1581, found 378.1576.

***N*-Trichloroethylcarbamate 28.** To a solution of **22** (100 mg, 0.22 mmol) in EtOH/pyridine (99:1, 5.5 mL) was added Pd black (17 mg), the suspension was degassed, and an H₂ atmosphere (1 atm) was applied. After being stirred at room temperature for 2 h, the suspension was filtered and washed with warm MeOH/pyridine (99:1), and the solution was concentrated and dried on the pump overnight. The crude material was dissolved in MeCN (6 mL) with Et₃N (90 μ L, 66 mg, 0.66 mmol), and the reaction was refluxed (95 °C) for 24 h. The solution was then cooled to 0 °C, and pyridine (23 μ L, 23 mg, 0.29 mmol) and TrocCl (56 mg, 36 μ L, 0.24 mmol) were added. After 3 h, LC–MS indicated the consumption of the amine, and the solvents were removed in vacuo. The brown residue was dissolved in 0.2 M HCl (5 mL), and the products were extracted with EtOAc (3 × 15 mL), dried (MgSO₄), filtered, and concentrated. The crude material was purified by flash chromatography (1:1, ethyl acetate/hexanes) to yield **28** (43 mg, 0.10 mmol, 45%) as a colorless solid. $R_f = 0.49$ (7:3, ethyl acetate/hexanes); mp >175 °C dec; $[\alpha]_D^{25} +5.5$ (c 1.0, CHCl₃); IR (neat) ν 3461, 1713; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 5.80 (d, $J = 3.2$, 0.5H), 4.78 (d, $J = 3.4$, 0.5H), 4.93 (d, $J = 11.9$, 0.5H), 4.83 (d, $J = 11.9$, 0.5H), 4.53–4.68 (m, 3H), 4.37 (d, $J = 3.3$, 0.5H), 4.35 (d, $J = 3.6$, 0.5H), 4.05–4.25 (m, 2H), 3.95 (d, $J = 2.2$, 0.5H), 3.92 (d, $J = 2.8$, 0.5H), 3.60–3.80 (m, 2H), 2.80 (br s, 0.4H), 2.40 (br s, 0.5H), 2.20 (m, 1H), 1.65 (s, 3H), 1.63 (m, 1H), 1.39 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 147.5, 146.9, 110.84, 110.79, 101.7, 93.2, 81.9, 81.8, 76.5, 74.7, 74.6, 73.6, 70.0, 69.9, 68.5, 66.8, 66.6, 54.1, 53.9, 41.0, 39.9, 31.1, 30.9, 29.9, 29.5; HRMS (ESI) calcd for C₁₅H₂₁-Cl₃NO₇ [M + H] 432.0378, found 432.0384.

Alcohol 29. To a solution of **28** (26 mg, 0.060 mmol) in dry CH₂Cl₂ (1 mL) was added Dess–Martin periodinane (39 mg, 0.092 mmol) at room temperature. After 2.5 h, a saturated solution of Na₂S₂O₃ (2 mL) was added, and the reaction was stirred for 20 min. The products were extracted with CH₂Cl₂ (3 × 3 mL), and the combined organics were washed with a saturated solution of NaHCO₃ (2 × 5 mL), dried (Na₂SO₄), filtered, and concentrated.

The residue was dissolved in CH₂Cl₂/MeOH (1:1, 1 mL) at room temperature, and to the stirred solution was added NaBH₄ (4 mg, 0.11 mmol). After 30 min, a saturated solution of NH₄Cl (2 mL) was added, and the reaction was stirred for 10 min. The layers were separated. The aqueous layer was extracted with EtOAc (3

× 4 mL), after which the organics were combined, dried (MgSO₄), and filtered, and the solvent was removed. The solid residue was purified by flash chromatography (1:1, ethyl acetate/hexanes) to yield **29** (23 mg, 0.053 mmol, 89%) as a colorless, glassy solid. $R_f = 0.50$ (2:3, ethyl acetate/hexanes); $[\alpha]_D^{25} -8.0$ (c 1.2, CHCl₃); IR (neat) ν 3460, 1613; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 5.82 (s, 0.4H), 5.81 (s, 0.6H), 4.9–5.05 (m, 1.4H), 4.85 (d, $J = 11.9$, 0.6H), 4.71 (d, $J = 11.9$, 0.6H), 4.63 (d, $J = 12.0$, 0.4H), 4.33 (d, $J = 3.0$, 0.6H), 4.28 (d, $J = 2.7$, 0.4H), 3.98–4.23 (m, 3.4H), 3.95 (d, $J = 2.4$, 0.4H), 3.78 (m, 2.4H), 3.59 (m, 0.4H), 3.43 (br s, 0.6H), 2.21 (m, 1H), 1.69 (m, 1H), 1.64 (s, 3H), 1.38 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 156.8, 114.1, 113.8, 105.5, 105.3, 94.8, 82.9, 81.4, 76.8, 75.2, 74.9, 73.0, 72.7, 67.9, 67.1, 66.4, 65.0, 52.9, 52.3, 38.3, 38.0, 27.9, 26.8, 26.1, 26.0, 25.7, 25.6; HRMS (ESI) calcd for C₁₆H₂₈NO₇S [M + H] 378.1581, found 378.1576.

***N*-tert-Butylsulfamate 31.** To a solution of **22** (200 mg, 0.44 mmol) in EtOH/pyridine (99:1, 10 mL) was added Pd black (25 mg), the suspension was degassed, and an H₂ atmosphere (1 atm) was applied. After being stirred at room temperature for 1.5 h, the suspension was filtered and washed with warm MeOH/pyridine (99:1), and the solution was concentrated and dried on the pump overnight.

The crude material was dissolved in MeCN (10 mL) with Et₃N (180 μ L, 133 mg, 1.31 mmol), and the reaction was refluxed overnight (95 °C). When LC–MS showed the disappearance of starting material, the reaction was cooled to room temperature, and Et₃N (310 μ L, 223 mg, 2.20 mmol) and then a 1.0 M solution of *tert*-butylsulfinyl chloride in CH₂Cl₂ (480 μ L, 0.48 mmol) were added. After 4 h, H₂O (10 mL) was added to the reaction, and the majority of the MeCN was removed in vacuo. The product was extracted with EtOAc (3 × 7 mL), dried (MgSO₄), filtered, and concentrated. The residue was filtered through silica to remove Et₃N and washed with EtOAc/MeOH (95:5). The solvent was removed in vacuo, and the residue was dissolved in CH₂Cl₂ (5 mL), to which *m*-CPBA (75 mg, 0.43 mmol) was added. After being stirred for 1.5 h at room temperature, a saturated solution of Na₂SO₃ (5 mL) was decanted into the reaction, and stirring was continued for a further 30 min. The layers were separated, and the organic layer was washed with a saturated solution of NaHCO₃ (2 × 5 mL) and dried (Na₂SO₄). After filtration and concentration, the crude material was purified by flash chromatography (7:3, ethyl acetate/hexanes) to yield **31** (85 mg, 0.23 mmol, 51%) as a colorless solid. $R_f = 0.54$ (ethyl acetate); mp > 175 °C dec; $[\alpha]_D^{25} -16.2$ (c 1.18, CHCl₃); IR (neat) ν 3482, 2254, 1314; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 5.80 (d, $J = 3.4$, 1H), 4.49 (d, $J = 3.3$, 1H), 4.30 (m, 4H), 3.95 (dd, $J = 13.1$, 3.0, 1H), 3.58 (m, 2H), 2.07 (br s, 1H), 1.86 (m, 1H), 1.65 (s, 3H), 1.63 (m, 1H), 1.43 (s, 9H), 1.38 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 113.6, 104.6, 81.6, 73.2, 70.8, 69.3, 68.6, 61.9, 56.3, 40.9, 27.6, 26.0, 25.7, 23.9; HRMS (ESI) calcd for C₁₆H₂₈NO₇S [M + H] 378.1581, found 378.1576.

***O*-Methyl Ether 33.** To a solution of **31** (121 mg, 0.32 mmol) in dry CH₂Cl₂ (6 mL) was added Dess–Martin periododinane (191 mg, 0.45 mmol) at room temperature. After 1 h, a saturated solution of Na₂S₂O₃ (6 mL) was added, and the reaction was stirred for 20 min. The products were extracted with CH₂Cl₂ (3 × 5 mL), and the combined organics were washed with a saturated solution of NaHCO₃ (2 × 6 mL), dried (Na₂SO₄), filtered, and concentrated.

The residue was dissolved in CH₂Cl₂/MeOH (1:1, 6 mL) at room temperature, and to the stirred solution was added NaBH₄ (19 mg, 0.33 mmol). After 30 min, a saturated solution of NH₄Cl (7 mL) was added, and the reaction was stirred for 10 min. The layers were separated. The aqueous layer was extracted with EtOAc (3 × 10 mL), after which the organics were combined, dried (MgSO₄), and filtered, and the solvent was removed.

The solid residue, **32**, was dissolved in anhydrous THF (6 mL), to which NaH (60% dispersion in mineral oil, 28 mg, 0.70 mmol) was added at room temperature. After being stirred for 20 min, MeI (56 μ L, 127 mg, 0.90 mmol) was added, and the reaction was stirred overnight. The reaction was then poured into a saturated

solution of NH₄Cl (6 mL), and the products were extracted with Et₂O (3 × 10 mL), which was dried (Na₂SO₄), filtered, and concentrated. The crude product was purified by flash chromatography (1:1, ethyl acetate/hexanes) to yield **33** (122 mg, 0.31 mmol, 95%, average of three runs) as a colorless foam. $R_f = 0.29$ (1:1, hexanes/ethyl acetate); $[\alpha]_D^{25} +6.5$ (c 1.19, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 5.85 (d, $J = 3.3$, 1H), 4.71 (br s, 1H), 4.20 (m, 2H), 3.89 (br s, 2H), 3.66 (br s, 3H), 3.53 (s, 3H), 1.88 (br s, 1H), 1.73 (m, 1H), 1.62 (s, 3H), 1.42 (s, 9H), 1.37 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 113.4, 105.4, 81.1, 75.1, 72.8, 71.2, 67.8, 61.6, 58.3, 51.5, 42.3, 27.7, 26.0, 25.6, 23.9; HRMS (ESI) calcd for C₁₇H₃₀NO₇S [M + H] 392.1738, found 392.1735.

Diphenyldithioacetal 34. Compound **33** (112 mg, 0.29 mmol) was dissolved into anhydrous CH₂Cl₂ (6 mL) under an Ar atmosphere, then PhSH (300 μ L, 318 mg, 2.90 mmol) was added, followed by Amberlyst-15 (280 mg), which was added in two portions in a 24-h interval. The reaction was stirred at room temperature for 60 h. After this time, a saturated solution of NaHCO₃ (8 mL) was poured into the reaction vessel and stirred for 30 min. The layers were separated, and the aqueous phase was extracted with CH₂Cl₂ (3 × 5 mL), dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by flash chromatography (65:35, hexanes/ethyl acetate) to yield **34** (129 mg, 0.23 mmol, 80%) as a colorless foam. $R_f = 0.41$ (1:1, hexanes/ethyl acetate); $[\alpha]_D^{25} +29.5$ (c 1.29, CHCl₃); IR (neat) ν 3451, 2250, 1307; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.44 (m, 4H), 7.28 (m, 6H), 5.00 (d, $J = 1.8$, 1H), 4.33 (m, 1H), 4.05 (d, $J = 1.7$, 1H), 3.95 (m, 3H), 3.67 (d, $J = 2.4$, 1H), 3.58 (m, 3H), 3.42 (s, 3H), 2.41 (ddd, $J = 14.5$, 13.1, 8.0, 1H), 2.10 (dd, $J = 14.6$, 4.3, 1H), 1.44 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 134.7, 134.1, 131.2, 130.8, 128.7, 128.6, 127.1, 127.0, 78.5, 75.2, 72.5, 67.3, 64.3, 61.8, 58.7, 57.6, 54.8, 42.7, 26.4, 24.0; HRMS (ESI) calcd for C₂₆H₃₅NO₆S₃Na [M + Na] 576.1519, found 576.1512.

Thioglycoside 35. A solution of **34** (125 mg, 0.23 mmol) in CH₂Cl₂ (10 mL) was cooled to 0 °C, and NBS (62 mg, 0.35 mmol) was added. After being stirred at 0 °C for 30 min, a saturated solution of Na₂S₂O₃ (15 mL) was added, and the biphasic mixture was stirred until colorless. The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 10 mL). The organics were combined, dried (Na₂SO₄), and filtered, and the solvents were evaporated in vacuo. The crude product was purified by flash chromatography (1:1, hexanes/ethyl acetate) to yield **35** (72 mg, 0.16 mmol, 71%) as a colorless foam. $R_f = 0.20$ (1:1, hexanes/ethyl acetate); $[\alpha]_D^{25} +170.8$ (c 0.72, CHCl₃); IR (neat) ν 3467, 2433, 1315; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.53 (dd, $J = 8.2$, 1.3, 2H), 7.28 (m, 3H), 5.75 (d, $J = 4.3$, 1H), 4.78 (m, 1H), 4.20 (dd, $J = 12.2$, 8.1, 1H), 4.08 (d, $J = 4.4$, 1H), 3.98 (m, 2H), 3.68 (br s, 3H), 3.53 (s, 3H), 2.92 (s, 1H), 1.83 (m, 2H), 1.42 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 135.2, 130.5, 128.7, 128.5, 126.7, 93.3, 76.9, 75.1, 74.4, 72.2, 70.5, 68.1, 58.3, 51.5, 42.1, 27.6, 23.9; HRMS (ESI) calcd for C₂₀H₃₀NO₆S₂ [M + H] 444.1509, found 444.1500.

***O*-Pivaloyl Ester 36.** To a solution of **35** (72 mg, 0.16 mmol) and DMAP (90 mg, 0.74 mmol) in anhydrous CH₂Cl₂/pyridine (2:1, 2 mL) was added PivCl (300 μ L, 294 mg, 2.44 mmol) under an Ar atmosphere at room temperature. After 5 h, the solvents were removed in vacuo (without heating), and the residue was dissolved in CH₂Cl₂ (6 mL), which was washed sequentially with 0.1 M HCl (2 × 3 mL) and H₂O (3 mL), dried (Na₂SO₄), filtered, and concentrated. The crude product was purified by flash chromatography (4:1, hexanes/ethyl acetate) to yield **36** (70 mg, 0.13 mmol, 81%) as a colorless foam and recrystallized by slow evaporation of CHCl₃. $R_f = 0.68$ (1:1, hexanes/ethyl acetate); $[\alpha]_D^{25} +118.2$ (c 0.45, CHCl₃); IR (neat) ν 2255, 1742, 1481, 1317; ¹H NMR (400 MHz, CDCl₃) δ (ppm) 7.54 (m, 2H), 7.30 (m, 3H), 5.84 (d, $J = 4.9$, 1H), 5.24 (d, $J = 4.3$, 1H), 4.76 (br s, 1H), 4.09 (m, 1H), 3.95 (m, 2H), 3.65 (m, 3H), 3.54 (s, 3H), 1.83 (m, 2H), 1.43 (s, 9H), 1.32 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) 176.3, 134.6,

130.8, 128.5, 127.0, 91.1, 75.2, 74.0, 72.0, 67.8, 65.5, 61.6, 58.2, 51.3, 42.1, 38.9, 26.8, 23.9, 14.9; LRMS (ESI) 528 (85%) [M + H]⁺.

***N*-Nucleoside 37.** A solution of **36** (24 mg, 0.046 mmol) in dry CH₂Cl₂ (0.7 mL) was added to a solution of bis-silylated *N*-acetylcytosine^{10b} (0.32 mmol of crude material) in dry CH₂Cl₂ (0.5 mL) under an Ar atmosphere at room temperature. NIS (41 mg, dried by lypophilization overnight) was added, followed immediately by triflic acid (5 μL, 8 mg, 0.055 mmol). Triflic acid (3 μL, 5 mg, 0.034 mmol) was added in two portions at 24-h intervals, and after 4 days the reaction was quenched with a saturated solution of Na₂S₂O₃ (3 mL) and a saturated solution of NaHCO₃ (2 mL) and stirred for 30 min. The products were extracted with EtOAc (4 × 6 mL), dried (MgSO₄), filtered, and concentrated. Flash chromatography of the crude material (4:1 to 1:4, hexanes/ethyl acetate) gave recovered starting material (10 mg, 0.019 mmol) and **37** (8 mg, 0.014 mmol, 52% based on recovered starting material) as a colorless solid. The crude product was recrystallized from EtOAc/hexanes. *R*_f = 0.32 (ethyl acetate); mp >210 °C dec; [α]²⁵_D +91.4 (*c* 0.35, EtOAc); ¹H NMR (400 MHz, CDCl₃) δ (ppm) 9.16 (br s, 1H), 8.53 (br s, 1H), 7.52 (d, *J* = 7.4, 1H), 5.86 (s, 1H), 4.90 (br s, 1H), 4.87 (s, 1H), 4.16 (dd, *J* = 12.1, 7.6, 1H), 4.05 (m, 1H), 3.82 (m, 3H), 3.60 (m, 1H), 3.48 (s, 3H), 2.25 (s, 3H), 1.83 (m, 1H), 1.52 (m, 1H), 1.46 (s, 9H), 1.27 (s, 9H); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) 177.4, 172.5, 164.0, 156.9, 144.9, 97.2, 93.4, 78.5, 75.0, 74.2, 71.6, 68.3, 67.8, 62.5, 58.4, 53.3, 39.2, 28.9, 27.0, 25.9, 24.1; LRMS (ESI) 571 (100%) [M + H]⁺.

Tricyclic *N*-Malayamycin A (7). The sulfonamide **37** (15 mg, 0.026 mmol) was dissolved in anhydrous CH₂Cl₂ (1.0 mL) under an Ar atmosphere. Anisole (43 μL, 43 mg, 0.39 mmol) and then triflic acid (8 μL, 14 mg, 0.10 mmol) were added, and the reaction was stirred at room temperature until LC–MS showed the disappearance of starting material. Trichloroacetyl isocyanate (13 μL,

20 mg, 1.04 mmol) and then anhydrous pyridine (7.5 μL, 7 mg, 0.093 mmol) were added, and the reaction was stirred at room temperature until LC–MS showed full conversion of the amine to the protected urea. At this point, the solvents were removed in vacuo (without heating), and the solid residue was stirred in 40 wt % MeNH₂ in H₂O/MeOH (3:1, 0.9 mL). After 24 h, 40 wt % MeNH₂ in H₂O solution (0.10 mL) was added, and the reaction was stirred for a further 48 h. The reaction was lypophilized, and the crude material was purified by preparative TLC (3:2, CHCl₃/MeOH) to yield **7** (5 mg, 0.014 mmol, 52%) as a colorless solid. *R*_f = 0.22 (75:25, CHCl₃/MeOH); [α]²⁵_D +25.8 (*c* 0.06, MeOH); ¹H NMR (400 MHz, D₂O) δ (ppm) 7.53 (d, *J* = 7.5, 1H), 5.88 (d, *J* = 7.6, 1H), 5.48 (s, 1H), 5.28 (br s, 1H), 4.07 (dd, *J* = 10.2, 5.6, 1H), 3.92 (d, *J* = 2.7, 1H), 3.79 (m, 3H), 3.55 (br s, 1H), 3.39 (br s, 1H), 3.32 (s, 3H), 1.79 (dd, *J* = 16.2, 6.1, 1H), 1.34 (m, 1H); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) 166.3, 161.0, 156.7, 139.4, 95.3, 94.4, 78.0, 75.3, 73.5, 71.1, 66.9, 56.6, 39.8, 36.3, 27.3; HRMS (ESI) calcd for C₁₅H₂₂N₃O₆ [M + H] 368.1565, found 368.1573.

Acknowledgment. We thank NSERC and Syngenta for generous financial support. We also thank Drs. Olivier Loiseleur (Syngenta, Basel, Switzerland) and Patrick Crowley (Syngenta, Jealott's Hill, U.K.) for stimulating discussions, Dr. Minh Tan Phan Viet for assistance with NMR experiments, and Dr. Michel Simard for X-ray crystal structure determination.

Supporting Information Available: ¹H NMR and ¹³C NMR spectra of compounds **9**, **11–19**, **21**, **22**, **24**, **26**, **28**, **29**, **31**, **33–37**, and **7** and X-ray crystallographic data for compounds **5**, **36**, and **37**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

JO061904R